

EURL-Salmonella Proficiency Test combined PPS-Food 2020

Detection of Salmonella in Hygiene swabs

Result Form

Laboratory information

Laboratory code	
Name contact person	
E-mail address contact person	
Name laboratory or institute	
Country	Country:
Did you participate as NRL PPS or NRL Food? For which <i>Salmonella</i> method(s) is your laboratory accredited?	NRL PPS NRL Food EN-ISO 6579-1:2017 EN-ISO 6579-1:2017/A1:2020 Other. Please specify:
Which method did you use in this PT?	EN-ISO 6579-1:2017 EN-ISO 6579-1:2017/A1:2020 Other namely:
Date and time of arrival of the parcel in your laboratory	dd/mm/yyyy 🏙 dd/mm/yyyy Time hh:mm
Was your parcel damaged at arrival?	No Yes, Description of damage:
Start date and time of storage at 5°C	dd/mm/yyyy
Start date of proficiency test	dd/mm/yyyy 🏥 dd/mm/yyyy

Positive control s	trol sample						
Which Salmonella serovar did you use in the control samples?	Enteritidis Typhimurium Goldcoast Nottingham Panama Poona Tranaroa (Salmonella enterica subsp. salamae) Other						
What was the concentration of the <i>Salmonella</i> control sample?	cfu/sample						
Was matrix added to the positive controle sample?	No Yes, specify the matrix						
Pre-enrichment - B	uffered Peptone Water (BPW)						
Temperature incubator BPW	°С						
Hours of incubation	hours						
Selective enrichment media							
Which selective enrichment media were used? (More options possible)	MKTTn (Mueller Kaufman Tetra Thionate-novobiocin broth) RVS (Rappaport-Vassiliadis medium with soda) MSRV (Modified Semi-solid Rappaport-Vasiliadis agar) Other, please specify						
MKTTn (Mueller Ka	aufman Tetra Thionate-novobiocin broth)						
Concentration novobiocin per 1L medium	mg/L						
Temperature incubation MKTTn							
pH at the day of use							

The state of the s				
RVS (Rappaport-Vassiliadis medium with Soya)				
Temperature incubation RVS °C				
pH at the day of use				
MSRV (Modified Semi-solid Rappaport-Vassiliadis agar) Concentration novobiocin per 1L medium Temperature incubation MSRV PH at the day of use				
pH at the day of use				
Selective Isolation media				
Which isolation media were used? (More options possible) Brilliant Green Agar (BGA) Brilliant Green Agar (BGA) Brilliant Green Agar (BSA) Brilliant Green Phenol-Red Lactose Sucrose (LBPLS) Brilliant Green Xylose Lysine Sulphonamide (BxLH) Rambach Salmonella Detection and Identification-2 (SM(ID)2) Xylose Lysine Deoxycholate (XLD) Other, please specify				
Confirmation of Salmonella suspected colonies				
What type of tests did you use for Serological Serotyping PCR Other Specify:				

Second Detection	Method (optional)	
Were the samples also tested with a second method?	Yes No	
What type of method? For example: PCR, qPCR, Rapid <i>Salmonella</i>		
Is this method validated?	Yes No	
Which organisation validated the method?		,
Please give a certificate number of reference for the validated method		
Is the second method used for analysing official control samples?	Yes No	
How many samples did you (approx.) analyse with this second method in 2019?		

Table 1: Confirmed Salmonella results

detected for positive confirmed result for Salmonella **not detected** for negative (confirmed) results for *Salmonella*

	EN- <u>ISO 6579-1</u>	<u>5000</u>	na metnoa
Samples ↓ B1	~	В1.	~
B2	~	B2.	~
B3	· ·	ВЗ.	~
В4	~	B4.	~
B5	· ·	B5.	~
B5		B5.	~
B6		B6.	~
B7		В7.	~
B8		B8.	×
B9	· ·	B9.	· ·
B10	· ·	B10.	· · · · · · · · · · · · · · · · · · ·
			· ·
B11		B11.	<u> </u>
B12	\ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \	B12.	×
B12		B12.	
B13	<u> </u>	B13.	
B14	~	B14.	
C1	<u> </u>	C1.	<u> </u>
C2	~	C2.	~
emarks and comments			
ame of person(s) arrying out the omparison study			
lame of the person in harge			

Original data is only accessible for EURL-Salmonella staff involved in this project.