

**Preview of the on-line Result form Cluster Analysis  
EURL-Salmonella Proficiency Test Typing 2019**



**EURL-Salmonella Proficiency Test Typing 2019**  
**Result form Cluster Analysis**

LABORATORY INFORMATION

Laboratory code PT 2019	<input type="text"/>
Name contact person (Cluster Analysis part)	<input type="text"/>
E-mail address contact person (Cluster Analysis part)	<input type="text"/>
Name laboratory or institute (Cluster Analysis part)	<input type="text"/>
Country	Country: <input type="text"/>


GENERAL

Did you serotype the strains?	<input type="radio"/> No <input type="radio"/> Yes
Strain SCA01 serovar name:	<input type="text"/>
Strain SCA02 serovar name:	<input type="text"/>
Strain SCA03 serovar name:	<input type="text"/>
Strain SCA04 serovar name:	<input type="text"/>
Strain SCA05 serovar name:	<input type="text"/>
Strain SCA06 serovar name:	<input type="text"/>
Strain SCA07 serovar name:	<input type="text"/>
Strain SCA08 serovar name:	<input type="text"/>
Strain SCA09 serovar name:	<input type="text"/>
Strain SCA10 serovar name:	<input type="text"/>

## REPORTING PFGE RESULTS

Do you want to submit PFGE results?  Yes  
 No

-> **Email the PFGE gel image as an uncompressed 8-bit gray scale TIFF file to [wilma.jacobs@rivm.nl](mailto:wilma.jacobs@rivm.nl).** Be sure to include your laboratory code in the name of the .tif file, preferably like: Lab01\_PFGE2019.tif

Date of emailing the PGFE gel image:   dd/mm/yyyy

-> Prepare the ZIP (Bionumerics 7) or XML export files (Bionumerics 6 or below), from the analysis in BioNumerics, including all test strains and reference strains, as well as the TIFF image. **Email these BN analysis data in a ZIP file to [wilma.jacobs@rivm.nl](mailto:wilma.jacobs@rivm.nl).** Be sure to rename your zip file to include your laboratory code in the name, preferably like: Lab01\_PFGE2019.zip

Date of emailing the BN analysis data:   dd/mm/yyyy

Which method did you use for PFGE?  Standard Pulsenet Protocol Salmonella PFGE  
 Standard Pulsenet Protocol Salmonella PFGE with modifications  
 Other:

Please enter the ID of the strains (SCA01 -SCA10, REF SB) in the corresponding position lanes on your gel (XbaI):

Lane 1	<input type="text"/>
Lane 2	<input type="text"/>
Lane 3	<input type="text"/>
Lane 4	<input type="text"/>
Lane 5	<input type="text"/>
Lane 6	<input type="text"/>
Lane 7	<input type="text"/>
Lane 8	<input type="text"/>
Lane 9	<input type="text"/>
Lane 10	<input type="text"/>
Lane 11	<input type="text"/>
Lane 12	<input type="text"/>
Lane 13	<input type="text"/>
Lane 14	<input type="text"/>
Lane 15	<input type="text"/>

How many clusters did you detect by PFGE data analysis?

0

1

2

3

Other

Please list the ID for the strains included in PFGE cluster 1

Please list the ID for the strains included in PFGE cluster 2

Please list the ID for the strains included in PFGE cluster 3

Please list the total number of bands per strain (> 33kb):

Strain SCA01

Strain SCA02

Strain SCA03

Strain SCA04

Strain SCA05

Strain SCA06

Strain SCA07

Strain SCA08

Strain SCA09

Strain SCA10

## REPORTING MLVA RESULTS

Do you want to submit MLVA results?

Yes

No

Please enter the reference to the MLVA scheme that was used:

Please list the loci in the scheme used

Locus 1:

Locus 2:

Locus 3:

Locus 4:

Locus 5:

Please list the allele profile per strain, using the format Locus 1-Locus 2-Locus 3-Locus 4-Locus 5 (expressed as e.g.: 02-09-07-03-02 or 03-12-10-00-211)

Strain SCA01	<input type="text"/>
Strain SCA02	<input type="text"/>
Strain SCA03	<input type="text"/>
Strain SCA04	<input type="text"/>
Strain SCA05	<input type="text"/>
Strain SCA06	<input type="text"/>
Strain SCA07	<input type="text"/>
Strain SCA08	<input type="text"/>
Strain SCA09	<input type="text"/>
Strain SCA10	<input type="text"/>

How many clusters did you detect by MLVA data analysis?

0  
 1  
 2  
 3  
 Other

Please list the ID for the strains included in MLVA cluster 1

Please list the ID for the strains included in MLVA cluster 2

Please list the ID for the strains included in MLVA cluster 3

## REPORTING WGS RESULTS

Do you want to submit WGS results?  Yes  No

-> **Transfer the raw reads** (fastq-files) to wilma.jacobs@rivm.nl, either by using wetransfer.com (multiple sessions may be required) or by uploading the files to the RIVM ftp server. Please contact wilma.jacobs@rivm.nl by email for further instructions on the use of the ftp server. Be sure to name your files to include your laboratory code and strain code in the name, preferably like: Lab01\_SCA01\_R1.fastq, Lab01\_SCA01\_R2.fastq, etc.

Date of sending the WGS fastq files:

-> **Email the distance matrix** (preferably as an .xls or .csv file) to wilma.jacobs@rivm.nl. Be sure to name the file to include your laboratory code, preferably like: Lab01\_Distance\_Matrix.xls

Date of emailing the distance matrix:

DNA extraction, library preparation and sequencing was performed:

In-house  
 Outsourced  
 Other:

WGS platform used:

Illumina Mi-Seq  
 Illumina NextSeq  
 Illumina HiSeq  
 Ion Torrent PGM  
 Ion Proton  
 Ion Torrent S5  
 PacBio  
 454  
 MinION  
 Other:

Please list (up to a maximum of 10) main **quantitative** criteria that were used to evaluate the quality of the sequence data, including the threshold used. (e.g. coverage, N50, number of contigs, etc.)

Quantitative criterium 1:

Threshold used for quantitative criterium 1:

Quantitative criterium 2:

Threshold used for quantitative criterium 2:

Quantitative criterium 3:

Threshold used for quantitative criterium 3:

Quantitative criterium 4:

Threshold used for quantitative criterium 4:

Quantitative criterium 5:

Threshold used for quantitative criterium 5:

Quantitative criterium 6:

Threshold used for quantitative criterium 6:

Quantitative criterium 7:

Threshold used for quantitative criterium 7:

Quantitative criterium 8:

Threshold used for quantitative criterium 8:

Quantitative criterium 9:

Threshold used for quantitative criterium 9:

Quantitative criterium 10:

Threshold used for quantitative criterium 10:

Please list (up to a maximum of 10) main **qualitative** criteria that were used to evaluate the quality of the sequence data, including the threshold if relevant. (e.g. contamination, confirmation of genus, etc.)

Qualitative criterium 1:

If relevant, threshold used for qualitative criterium 1:

Qualitative criterium 2:

If relevant, threshold used for qualitative criterium 2:

Qualitative criterium 3:

If relevant, threshold used for qualitative criterium 3:

Qualitative criterium 4:

If relevant, threshold used for qualitative criterium 4:

Qualitative criterium 5:

If relevant, threshold used for qualitative criterium 5:

Qualitative criterium 6:

If relevant, threshold used for qualitative criterium 6:

Qualitative criterium 7:

If relevant, threshold used for qualitative criterium 7:

Qualitative criterium 8:

If relevant, threshold used for qualitative criterium 8:

Qualitative criterium 9:

If relevant, threshold used for qualitative criterium 9:

Qualitative criterium 10:

If relevant, threshold used for qualitative criterium 10:

Please select the analysis used for the WGS data

- SNP-based - reference-based
- SNP-based - assembly-based
- cg-MLST-based
- wg-MLST-based
- Other:

Please select the tools used for analysis:

- Applied Math
- BioNumerics
- Enterobase
- Ridom SeqSphere
- Other:

Which method did you use for phylogenetic analysis?

- Maximum likelihood (ML)
- Neighbor joining (NJ)
- Bayesian
- Other:

How many clusters did you detect by WGS data analysis?

- 0
- 1
- 2
- 3
- Other:

Please list the ID for the strains included in WGS cluster 1

Please list the ID for the strains included in WGS cluster 2

Please list the ID for the strains included in WGS cluster 3

**FINALLY**

Any comments: