



Protocol EURL-*Salmonella* Proficiency Test Typing 2019

1. Introduction

The European Union Reference Laboratory (EURL) - *Salmonella* organises the 24th Proficiency Test (PT) on typing of *Salmonella* strains amongst the National Reference Laboratories for *Salmonella* (NRLs-*Salmonella*).

The main objective of this typing PT is to test the performance of the participating laboratories for serotyping of *Salmonella* spp.

A cluster analysis part, using PFGE and/or MLVA and/or WGS data, is optionally available as a pilot study.

The PT will take place in week 45 and onwards. The timetable can be found on page 5 of this protocol.

All data have to be reported through an electronic result form. The link to this form will be sent by email in week 45. **Submission of serotyping data** has to be finalised on **13 December 2019** at the latest.

The part on cluster analysis will use a separate result form, and the link to this form will be sent to the participants in a separate email in week 45 as well. Deadline for the electronic **submission of all cluster analysis results** is **31 January 2020** at the latest.

Note that in 2018 the EURL-Salmonella changed to a new system (formdesk) for the web-based registration and result forms. A short guidance to use this system for data submission will be included in the emails sending the links.

2. Transportation of the *Salmonella* strains to the laboratories

The strains for the serotyping part and optionally the cluster analysis part of the study will be transported all in one (larger) parcel. The strains will be sent as Biological Substance Category B (UN 3373) with a door-to-door courier to your laboratory.

The shipment of the strains is scheduled for Monday 4 November 2019.

3. Serotyping

A total number of 20 *Salmonella* strains (coded S1 - S20) have to be serotyped. An additional *Salmonella* strain (S-21), being a less common *Salmonella* serovar, is also included in the package and serotyping of this strain is optional.

The method routinely performed in your laboratory has to be used in the study.

Each laboratory is allowed to send strains for serotyping to another reference laboratory in their country, if this is part of the normal routine procedure.

As explained at the recent EURL-*Salmonella* Workshops, please note to be very careful in following the exact instructions of the various manufacturers of the different antisera available.

The results for each strain have to be reported with the formula for the O-antigens and H-antigens and the serovar names according to the White-Kauffman-le Minor scheme of 2007

(https://www.pasteur.fr/sites/default/files/veng_0.pdf)

Laboratories have to report only those results, on which the identification of serovar names is based.



If, based on the results as obtained, a definite conclusion on the serovar name cannot be given, then identify the strains by giving the antigenic formula as far as detected.

Examples of preferred reporting:

O-antigens	H-antigens (phase 1)	H-antigens (phase 2)	Serovar name
9,12	g,m	-	Enteritidis
4,12	i	2	Typhimurium
4,5,12	i	-	4,5,12:i:-
4	i	-	4:i:-
6,7	-	1,5	6,7:-:1,5
42	g,t	-	42:g,t:-

The evaluation of the serotyping results will be performed by the EURL-*Salmonella* according to Table 1.

Table 1. Evaluation of serotyping results

Results	Evaluation
Auto-agglutination or Incomplete set of antisera (outside range of antisera)	Not typable
Partly typable due to incomplete set of antisera or Part of the formula (for the name of the serovar) or No name serovar	Partly correct
Wrong serovar or mixed sera formula	Incorrect

Hendriksen et al. (J Clin Microbiol 47(9): 2729-2736) reported that colonial form variation may occur with the expression of the O:6₁ antigen by some serogroup C₂ serovars.

Concerning the EURL-*Salmonella* PTs on serotyping it was decided to consider the serovar pairs involved (e.g. *S. Newport/S. Bardo* and *S. Hadar/S. Istanbul*) not as distinct serovars, though they should be reported as actually typed by the participants. Nevertheless, typing should include testing for the presence of O:6 antigen.

In practice this means that for example a 6,8:z₁₀:e,n,x typed strain has to be reported as Hadar, and a 8:z₁₀:e,n,x typed strain has to be reported as Istanbul, but that either result is considered as correct.

In 2007, criteria for 'good performance' have been defined (Mooijman, 2007). Penalty points are given for strains that are typed incorrectly. A distinction is made between the five most important human health related *Salmonella* serovars (as indicated in EU legislation) and all other strains:

- **4 penalty points:** incorrect typing of *S. Enteritidis*, *S. Typhimurium* (including the monophasic variant), *S. Hadar*, *S. Infantis* or *S. Virchow* **or** assigning the name of one of these five serovars to another strain.
- **1 penalty point:** incorrect typing of all other *Salmonella* serovars.



The total number of penalty points is calculated for each NRL-*Salmonella*. The criterion for good performance is set at less than four penalty points. All EU Member State NRLs not meeting the criterion of good performance (results with four penalty points or more) have to participate in a follow-up study.

4. Pilot on cluster analysis

The cluster analysis part of the PT typing is optional. It includes 10 *Salmonella* strains (coded SCA01 – SCA10) and can be performed up to the choice of the participant by PFGE and/or MLVA and/or WGS (or any combination of these methods). Participants are asked to test these strains using their own routine method(s) of choice. Details on the method(s) used and the outcome of the cluster analysis have to be reported in the **electronic result form**. Additionally, specific data for PFGE or WGS have to be send by email or have to be uploaded to a ftp server. Detailed instructions for this will be sent to the participants in week 45, along with the link to the result form.

Results to be submitted are listed below.

PFGE:

- **Electronic result form:** protocol used, position of the lanes, total number and shared number of bands per profile, cluster identification
- **Email the PFGE gel image** as an *uncompressed 8-bit gray scale TIFF file* to wilma.jacobs@rivm.nl . Be sure to include your laboratory code in the name of the .tif file, preferably like: Lab01_PFGE2019.tif
- Prepare the ZIP (Bionumerics 7) or XML export files (Bionumerics 6 or below), from the analysis in BioNumerics, *including all test strains and reference strains, as well as the TIFF image*. **Email these BN analysis data in a ZIP file** to wilma.jacobs@rivm.nl . Be sure to rename your zip file to include your laboratory code in the name, preferably like: Lab01_PFGE2019.zip

MLVA:

- **Electronic result form:** scheme/loci used, the allelic profile, cluster identification

WGS:

- **Electronic result form:** background information on the wet-lab and dry-lab methods used, cluster identification (SNP-based or cgMLST/wgMLST-based)
- **Upload the raw reads** (fastq-files) to the ftp server according to the instructions to be sent in week 45. Be sure to name your files to include your laboratory code and strain code in the name, preferably like: Lab01_SCA01_R1.fastq, Lab01_SCA01_R2.fastq, etc.

Evaluation (per methodology) of the participants' cluster analysis results will be done on the ability to correctly identify cluster(s) of genetically closely related isolates, as pre-defined by the EURL-*Salmonella*.



5. Reporting of the PT results

As usual, all data have to be reported through an electronic result form. The link to the serotyping result form will be sent by email to the participants in week 45. **Submission of serotyping data** has to be finalised on **13 December 2019** at the latest.

The part on cluster analysis will use a separate result form, and the link to this form will be sent to the participants in a separate email in week 45 as well. Deadline for the electronic **submission of all cluster analysis results** (result form plus additional PFGE and/or WGS data as requested) is **31 January 2020** at the latest.

Mind that the electronic result forms are no longer accessible after these deadlines!

In case you foresee problems with the deadline(s), please contact us beforehand.

If you have questions or remarks about this study, or in case having problems using the electronic result forms, please contact:

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**Timetable EURL-*Salmonella* Proficiency Test Serotyping and
optional Proficiency Test PFGE and/or MLVA and/or WGS Cluster Analysis
- 2019**

Week	Date	Topic
39	23 September	Emailing of the link to the registration form for the typing study. Please register by 18 October at the latest.
43	21-25 October	Emailing of the protocol 2019.
45	4-8 November	Shipment of the parcels to the participants as Biological Substance Category B (UN 3373). If you did not receive the parcel by 8 November , please contact the EURL- <i>Salmonella</i> .
45	4-8 November	Sending the link for the result form on serotyping to the participants.
45	4-8 November	Sending the link for the result form on PFGE and/or MLVA and/or WGS Cluster Analysis to the participants in a separate email.
45	4-8 November	<i>Upon receipt</i> : Starting the identification of the strains, according to the usual practice of the laboratory.
50	13 December 2019 at the latest	Deadline for completing the electronic submission of serotyping results: 13 December 2019 After this deadline, the result form for serotyping will be closed.
	31 January 2020 at the latest	Deadline for completing the electronic submission of PFGE/MLVA/WGS Cluster Analysis results: 31 January 2020
	February 2020	Serotyping: Evaluation of individual laboratory results and Interim Summary Report.
	April/May 2020	Pilot PFGE/MLVA/WGS Cluster Analysis: Evaluation of individual laboratory results and Summary Report.
	Summer 2020	Final report.