



National Institute for Public Health  
and the Environment  
*Ministry of Health, Welfare and Sport*

# Whole genome sequence comparison of MDR *Salmonella* Infantis isolates from broilers and humans in the Netherlands

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## Introduction

- In recent years a shift is seen in the most prevalent *Salmonella* serotypes in broilers in the Netherlands
  - ❖ *Salmonella* Paratyphi B variant Java was the most prevalent serotype
  - ❖ Increase in the relative percentage of *Salmonella* Infantis.
- In 2018 *S. Infantis* was the most reported serotype in broilers as well as poultry meat in the Netherlands as well as in Europa
- *S. Infantis* ranked fourth among the serotypes isolated from human salmonellosis cases in the Netherlands and in Europe in the past few years
- *S. Infantis* increasingly associated with multi-drug resistance.
- In 2014, a multi-drug resistant *S. Infantis* strain was identified carrying a mega-plasmid of approximately 300 Kb called pESI (plasmid of emerging *S. Infantis*)



## Introduction

- Yearly every EU member state has to report about prevalence and trends of zoonotic pathogens in animals, animal products and humans to ECDC and EFSA (zoonoses Directive (2003/99/EC)).
- Information regarding prevalence and risk factors of zoonotic pathogens in animals is required for adequate prevention measures.

The Netherlands: NVWA/RIVM project surveillance livestock

- NVWA (Netherlands Food and Consumer Product Safety Authority);
  - ❖ 'Monitoring pathogens in livestock'
- RIVM (National Institute of Public Health and the Environment);
  - ❖ 'Monitoring of gastro-enteritis (GE) and GE-pathogens in humans'



## Introduction

- Trends in prevalence of zoonotic pathogens in different livestock sectors:
  - ❖ Pigs (2013)
  - ❖ Layers (2015)
  - ❖ Beef cattle (2017)
  - ❖ Broilers (2018-2019)
  - ❖ Dairy cattle (2020)
  - ❖ Small ruminants; Dairy Goats/Sheep (2016)
- Research in livestock & farmers, family members and employees
- Faecal sampling and questionnaire



## Broilers (2018-2019)

### Zoonotic pathogens:

- Animals: *Campylobacter*, ESBL/AmpC-producing *E. coli*, *Listeria monocytogenes*, *Salmonella* and STEC.
- Humans: *Campylobacter*, ESBL/AmpC-producing *E. coli*, MRSA and *Salmonella*
  - ❖ The Netherlands: 625 broiler farms on 823 locations (CBS, 2018)
  - ❖ Randomly selected farms with >3000 broilers (n=230)
  - ❖ 200 put in a sampling scheme, 30 were backups

### Response

- 198 farms visited; 379 flocks tested
- 81 farms (40.6%) participated in the human study; 132 farmers, family members and employees



## Broilers (2018-2019)

Due to

- Commission Regulation (EU) No 200/2010 (reduction of the prevalence of *Salmonella* serotypes in adult breeding flocks of *Gallus gallus*) implementing Regulation (EC) No 2160/2003 (on the control of *Salmonella* and other specified food-borne zoonotic agents);

The yearly maximum percentage of adult breeding flocks of *Gallus gallus* remaining positive for the relevant\* *Salmonella* serotypes is to be 1% or less

- Wait for results of participating farms from the National Control Program *Salmonella* (AVINED)
- After the end of the complete sampling period *Salmonella* isolation was performed from the stored broiler faeces samples (1:1 glycerol, -20 °C)



## Methods *Salmonella* (1)

### ❖ Animals

- 1 or 2 Broiler houses sampled
- Three faecal samples (12 droppings each) collected (front, middle and back)
- 10g Faeces/glycerol of each sample of was pooled to 30 g faeces/glycerol per broiler house
- Incubated overnight together with 270 ml BPW at 37 °C

### ❖ Humans

- 1 g Faeces was added to 9ml BPW and incubated overnight at 37 °C



## Methods *Salmonella* (2)

- 100 µl Transferred to a MRSV agar plate supplemented with novobiocin and incubated at 41.5 °C for 24 and 28 h
- Suspected plates were streaked on BSA plates and incubated at 37 °C (18-22h)
- Biochemical confirmation
- Obtained isolates were analysed with a xMap *Salmonella* Serotyping kit (Biovet)
  
- *S. Infantis* isolates were analysed with short-read sequencing on a NovaSeq (Illumina) platforms.
- Core genome MLST (cgMLST) analysis was performed after *de novo* assembly.
- Isolates were screened for pESI plasmid linked genes as well as antibiotic resistance (AR) genes and *Salmonella* virulence factors.





## Results *Salmonella*\_broilers

	# Farms (houses)	Positive farms (houses)	Prevalence
RIVM	194 (371)	23 (35)	11.9% (9.4%)

Serotype	# Farms	Prevalence	# houses
S. Agona	1	4.3%	1
S. Infantis	10	43.5%	14
S. Paratyphi B variant Java	12	52.2%	20

- Farm-level prevalence of 5.1% and flock-level prevalence of 3.7% for *S. Infantis*



## Results *Salmonella*\_humans

- Response rate; 40.6%
  - ❖ 132 farmers, family members and employees
  - ❖ 81 farms
  
- Prevalence; 0.8%
  - ❖ 1 farmer positive with *S. Infantis*, no complaints
  - ❖ Flocks were *Salmonella* negative at time of sampling



## Whole Genome Sequencing

- Broiler isolates (14)
- Human isolate; farmer (1)
  
- Routine *Salmonella* surveillance at RIVM (Centre of Diagnostics and Laboratory Surveillance (IDS))
  - ❖ >100 *S. Infantis* isolates (July 2018 – May 2019)
  - ❖ Focus on MDR isolates (phenotypic data)
  
- Human isolates (13), Pig (2), Broiler (1), Broiler meat product (1)



- All 32 *S. Infantis* isolates belonged to ST32

Cluster 2





## Virulence Factors (VFs)

➤ 233 *Salmonella* VFs from public databases and literature

➤ Listed in  **frontiers**  
in Microbiology

ORIGINAL RESEARCH  
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### Phenotypic Prediction: Linking *in vitro* Virulence to the Genomics of 59 *Salmonella enterica* Strains

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➤ Implemented in one target scheme (SeqSphere<sup>+</sup>)



## VFs

- 156 VFs present in all *S. Infantis* isolates investigated
- Most of these VF gene sequences were identical (83.3%; 130/156)
- Ten genes contained a synonymous SNP which does not result in an amino acid substitution



## VF<sub>s</sub>

Gene	Function	SNP	AA substitution	Source (#)
<i>fimD</i>	Outer membrane usher protein	C1855A	R619S	Pig (1)
<i>fimZ</i>	Fimbria biosynthesis transcriptional regulator	C136T	R46C	Pig (1)
<i>fimF</i>	Type I fimbriae adaptor protein	T62C	V21A	Broiler (1)
<i>lpfC</i>	Long polar fimbrial usher protein	2451 INS 1nt	PMSC	Human (3)
<i>stfA</i>	Fimbrial subunit	G192A	M64I	Broiler (2)
<i>tcfC</i>	Fimbrial outer membrane usher	G1201A	M67I	Broiler (2)
<i>mgtB</i>	Magnesium-translocating P-type ATPase	T1662A	D554E	Human (1)
<i>ratB</i>	Outer membrane protein	T3898C	Y1300H	Broiler (2)
<i>sinI</i>	Outer membrane protein	T589G	E197PMSC	Broiler (13), Human (6), Farmer (1)
<i>hilC</i>	Transcriptional regulator	T310C	Y104H	Human (1)
<i>ssrA</i>	Sensor kinase	C1894T	H632Y	Broiler (1)
<i>sscB</i>	Secretion system chaperone	T238A	F80I	Broiler (13), Human (6), Farmer (1)
<i>ssaU</i>	Type III secretion system protein	G353A	S118N	Pig (1)
<i>slrP</i>	E3 ubiquitin-protein ligase	C178A	A60I	Broiler (2)
<i>sopA</i>	Type III secretion system effector	T629A	I210K	Human (1)
<i>sifB</i>	Type III secretion system effector	G766A	V256I	Broiler (2)
<i>sspH2</i>	Type III secretion system effector	G1224T	M408I	Broiler (1)

PMSC = premature stop codon



## *In silico* analysis of AMR genes and point mutations in *gyrA* and *parC*

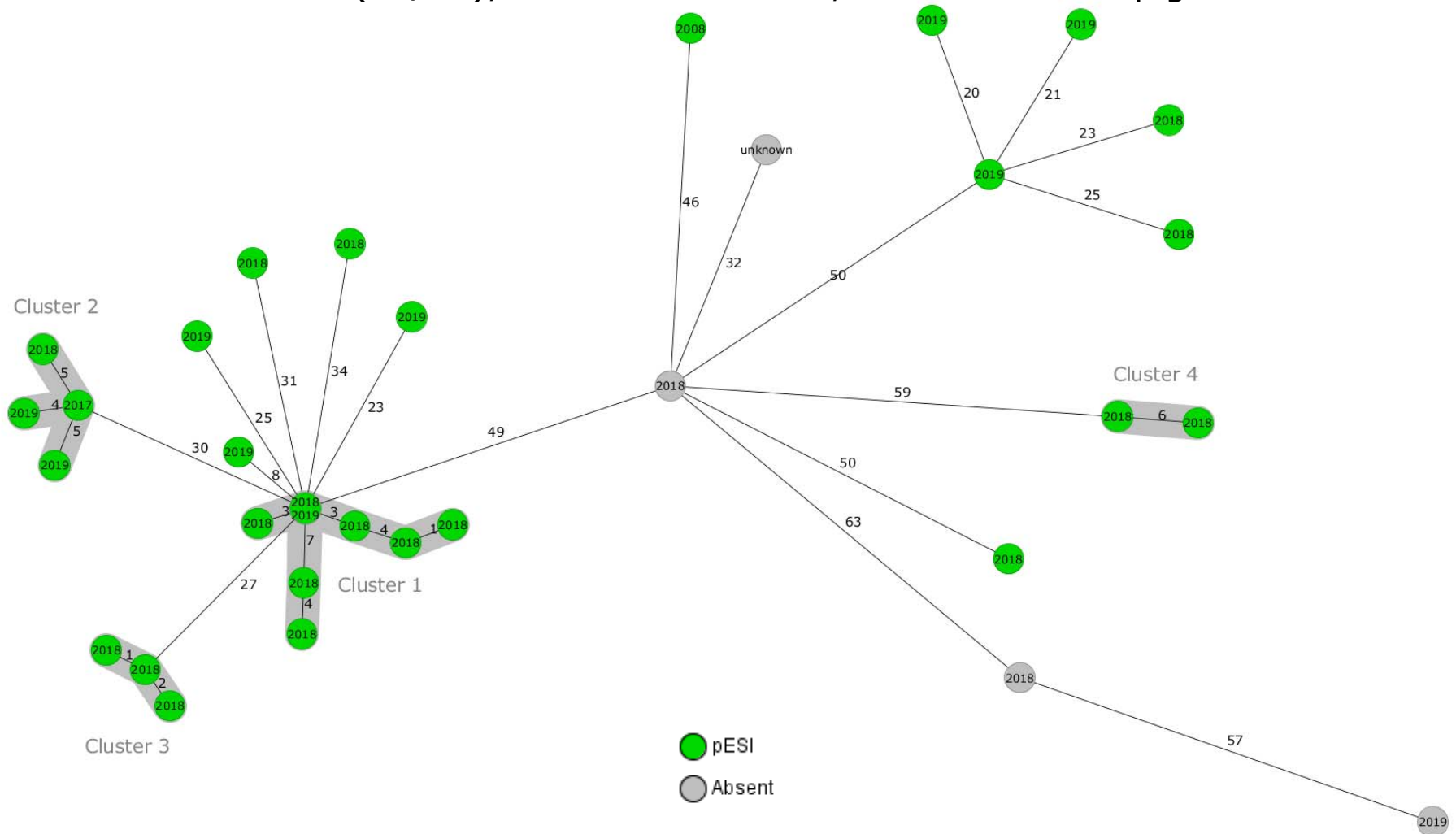
Source	#	<i>aadA</i>	<i>aph(3')-I</i>	<i>aph(4)-I</i>	<i>str</i>	<i>bla</i>	<i>sul</i>	<i>tet</i>	<i>dfr</i>	GyrA p83*	GyrA p87*
Farm P	2	<i>aadA1</i>					<i>sul1</i>	<i>tet(A)</i>	<i>dfrA14</i>	WT	<b>G</b>
Farm Q, R, S, T, U,V	8	<i>aadA1</i>	<i>aph(3')-Ic</i>				<i>sul1</i>	<i>tet(A)</i>	<i>dfrA14</i>	WT	<b>G</b>
Farm W	2	<i>aadA1</i>					<i>sul1</i>	<i>tet(A)</i>		<b>Y</b>	WT
Farm X	1	<i>aadA1</i>					<i>sul1</i>	<i>tet(A)</i>		WT	<b>G</b>
Farm Y	1									WT	WT
Farm Z, farmer	1	<i>aadA1</i>					<i>sul1</i>	<i>tet(A)</i>	<i>dfrA14</i>	WT	<b>G</b>
Broiler	1	<i>aadA1</i>	<i>aph(3')-Ic</i>				<i>sul1</i>	<i>tet(A)</i>	<i>dfrA14</i>	WT	<b>G</b>
Broiler meat	1	<i>aadA1</i>	<i>aph(3')-Ic</i>		<i>strAB</i>	TEM-1b	<i>sul1, sul2</i>	<i>tet(A)</i>	<i>dfrA8</i>	WT	<b>G</b>
Pig	1				<i>strAB</i>		<i>sul1, sul2</i>	<i>tet(A)</i>	<i>dfrA23</i>	WT	WT
	1									WT	WT
Human	3	<i>aadA1</i>	<i>aph(3')-Ic</i>			CTX-M-65	<i>sul1</i>	<i>tet(A)</i>	<i>dfrA14</i>	WT	<b>Y</b>
	3	<i>aadA1</i>					<i>sul1</i>	<i>tet(A)</i>	<i>dfrA14</i>	WT	<b>G</b>
	1	<i>aadA1</i>	<i>aph(3')-Ic</i>			TEM-1b	<i>sul1</i>	<i>tet(A)</i>	<i>dfrA14</i>	WT	<b>G</b>
	1	<i>aadA1</i>	<i>aph(3')-Ic</i>				<i>sul1</i>	<i>tet(A)</i>		WT	<b>G</b>
	1	<i>aadA1</i>	<i>aph(3')-Ic</i>	<i>aph(4)-Ia</i>		CTX-M-65	<i>sul1</i>	<i>tet(A)</i>	<i>dfrA14</i>	WT	<b>Y</b>
	1	<i>aadA1</i>				CTX-M-65	<i>sul1</i>	<i>tet(A)</i>	<i>dfrA14</i>	WT	<b>Y</b>
	1	<i>aadA1</i>					<i>sul1</i>	<i>tet(A)</i>		WT	<b>G</b>
	1		<i>aph(3')-Ia</i>		<i>strAB</i>		<i>sul2</i>	<i>tet(A)</i>	<i>dfrA32</i>	<b>Y</b>	WT
	1									WT	WT





pESI

- Prevalence 87.5% (28/32), absent in 1 broiler, 1 human and 2 pig isolates



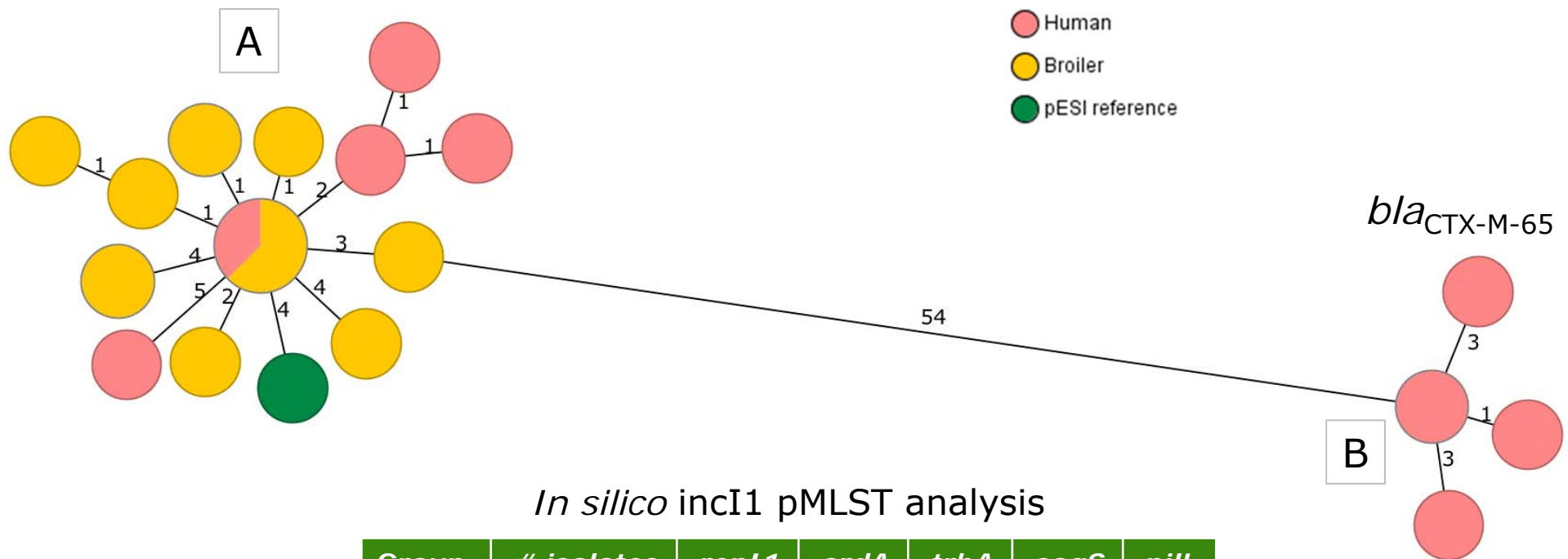


## Whole plasmid MLST (wp(MLST))

- Reference; *Salmonella enterica* subsp. *enterica* serovar Infantis strain 119944 plasmid pESI, complete sequence (CP047882 (285,081 bp))
  
- A wpMLST scheme was constructed containing 282 gene targets (SeqSphere<sup>+</sup>)
  - ❖ IncI1-type genes (*traE*, *traF...traX*, *trbA...trbC*)
  - ❖ IncP replicase
  - ❖ Plasmid SOS inhibitors (*psiA*, *psiB*)
  - ❖ Fimbrial operon (*faeA*, *faeD*, *faeH*)
  - ❖ Type IV pilus operon (*pilL..pilR*, *pilX*)
  - ❖ Class 1 integron (*intI1*), class 2 integrons (*intI2*)
  - ❖ AMR genes (*aadA1*, *dfrA14*, *sul1*, *tet(A)*)
  - ❖ Mercury resistance operon (*merCBDEPTR*)
  - ❖ Yersiniabactin operon (*fyuA*, *ybtETU*, *irp1*, *irp2*, *ybtAAPQXS*)



## pESI wpMLST



*In silico* incI1 pMLST analysis

Group	# isolates	<i>repI1</i>	<i>ardA</i>	<i>trbA</i>	<i>sogS</i>	<i>piL</i>
A	22	NA	2	21	9	3
	1	NA	2	46	9	3
B	5	NA	11	8	14	3



## Conclusions (1)

- Surveillance farm animals (broilers)
  - ❖ 198 farms visited; 379 flocks tested
  - ❖ Participation in the human study; 40.6% (81 farms, 132 persons)
  - ❖ *Salmonella* prevalence on the farms; 11.9%, the flocks; 9.4%
  - ❖ *S. Infantis* prevalence farm-level; 5.1%, flock-level; 3.7%
  - ❖ *Salmonella* prevalence in the human study; 0.8%
  - ❖ Human isolate; *S. Infantis*



## Conclusions (2)

### ➤ WGS analysis

- ❖ All belonged to ST32
- ❖ cgMLST analysis revealed considerable variation among the isolates included in this study. Even isolates from different flocks of the same farm were never 100% identical, although they did cluster together
- ❖ 156 VFs present in all *S. Infantis* isolates investigated
- ❖ Most of these VF gene sequences were identical (83.3%)
- ❖ Sixteen genes contained a non-synonymous SNP which resulted in an amino acid substitution or PMSC



## Conclusions (3)

### ➤ WGS analysis

- ❖ *In silico* screening of the assembled genomes showed a high prevalence of 5 antibiotic resistance genes (*aadA1*, *aph(3')*-Ic, *drfA14*, *sul1*, *tet(A)*)
- ❖ Five human *Infantis* isolates were ESBL producers (*bla*<sub>CTX-M-65</sub>)
- ❖ (Fluoro)quinolone resistance was prevalent among the *Infantis* isolates due to mutations in *gyrA*, which resulted in amino acid substitutions
- ❖ pESI prevalence was 87.5% (28/32)
- ❖ Based on (w)pMLST two subtypes of pESI were shown



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